AGRICULTURAL AND FOOD CHEMISTRY

Design, Synthesis, and Particular Biological Behaviors of Chain-Opening Nitromethylene Neonicotinoids with Cis Configuration

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ABSTRACT: On the basis of the structure of heterocyclic-fused cis configuration derivatives and chain-opening neonicotinoids, two series of novel chain-opening tetrahydropyridine analogues were designed and synthesized. The preliminary bioassay tests were determined on cowpea aphid (*Aphis craccivora*) and armyworm (*Pseudaletia separata* Walker). The results showed that some of the target compounds exhibited repellent effects, whereas others showed good insecticidal activities.

KEYWORDS: chain-opening, neonicotinoids, repellent effects, cis configuration, tetrahydropyridine

INTRODUCTION

Neonicotinoids, as the fourth-generation insecticides after organophosphorus, carbamates, and pyrethroids, are the fastest growing synthetic insecticides on the market.¹ Their global sales reached U.S. \$2.632 billion in 2009 and accounted for 23.7% of total market share of insecticides in 2008,² which were expected to take the biggest market of insecticides. With a thorough comprehension of the action mechanism, neonicotinoids are a class of insecticides targeting the insect nicotinic acetylcholine receptors (nAChRs).^{3,4}

There are four common molecular features of neonicotinoids: (1) an aromatic heterocycle, (2) flexible linkage, (3) hydroheterocycles or guanidine/amidine, and (4) an electronwithdrawing group.⁵ According to the structural characteristics of the third feature, the commercialized neonicotinoids can be subdivided into two categories: (1) cyclic compounds, including imidacloprid 1 (Figure 1), thiacloprid 2 (Figure 1), and thiamethoxam 3 (Figure 1); and (2) acyclic neonicotinoids, including nitenpyram 4 (Figure 1), acetamiprid 5 (Figure 1), clothianidin 6 (Figure 1), dinotefuran 7 (Figure 1), and sulfoxaflor 8 (Figure 1). Generally, compared with cyclic ones, acyclic neonicotinoids, which are defined as ring-opening or chain-opening neonicotinoids, exhibit a broader insecticidal spectrum.^{6,7} The acyclic diamine skeleton was considered to be an effective bioisostere of imidazolidine and other heterocycles,⁸ which made it possible for the reconstruction of an acyclic bioisostere to often acquire desirable bioactivities .

In previous work, we had reported the synthesis, bioactivities, and mode of action of nitromethylene neonicotinoids with fixed cis configuration.⁹ The nitro group was fixed to form the cis configuration through the introduction of a tetrahydropyridine ring, which helps to improve the photostability. The preliminary bioassay results showed that their insecticidal activities against resistant strain brown planthopper were higher than that of imidacloprid.⁵ Accordingly, it attracted us to investigate what chain-opening analogues of cis configuration nitromethylene neonicotinoids would provide.

To further explore the bioactivities of chain-opening analogues of cis configuration nitromethylene neonicotinoids, two series of compounds with tetrahydropyridine-fused neonicotinoids were designed and synthesized. Series 1 (see Table 1) was designed and synthesized on the basis of the structures of nitenpyram 4 (Figure 2) and acetamiprid 5 (Figure 2), wherein the substituent R_3 was an alkyl group. In the design of series 2, R_3 was replaced by a hydrogen atom as found in the structures of clothianidin 6 (Figure 2) and dinotefuran 7 (Figure 2).

In this paper, the syntheses of two series of chain-opening nitromethylene neonicotinoids and their particular biological behaviors or bioactivities are reported.

MATERIALS AND METHODS

Instrumentation and Chemicals. Unless otherwise noted, reagents and solvents were used as received from commercial suppliers. Melting points (mp) were recorded on a Büchi B540 apparatus and are uncorrected. ¹H NMR spectra were recorded on a Bruker WP-500SY (500 MHz) or Bruker WP-400SY (400 MHz) spectrometer with CDCl₃ as the solvent and TMS as the internal standard. Chemical shifts are reported in δ (parts per million) values. High-resolution mass spectra were recorded under electron impact (70 eV) conditions using a MicroMass GCT CA 055 instrument. Combustion analyses for elemental composition were made with an Elementar vario EL III. GC mass spectra were recorded using a HP 6890 gas chromatograph and HP 5973 mass selective detector. Analytical thin-layer chromatography (TLC) was carried out on precoated plates (silica gel 60 F254), and spots were visualized with ultraviolet (UV) light.

Synthetic Procedures. Synthesis of Series 1 (Scheme 1). N-((6-Chloropyridin-3-yl)methyl)ethanamine (19). Compound 19 was prepared as described in the literature¹⁰ in 70.3% yield: GC-MS, m/z (%) 170 ([M]⁺, 20), 155 (80), 126 (100), 114 (10), 90 (12).

N-((6-Chloropyridin-3-yl)methyl)-N-ethyl-1-(methylthio)-2-nitroethenamine (20). Compound 20 was prepared as described in the *literature*¹¹ in 18.5% yield: GC-MS, *m/z* (%) 242 ([M]⁺ – 46, 53), 227 (15), 213 (100), 169 (45), 155 (28), 141 (29), 126 (91), 90 (12).

N-Methyl-1-(methylthio)-2-nitroethenamine (22). To the solution of compound 21 (15.0 g, 90 mmol) in acetonitrile (50 mL) was added dropwise 60–75% aqueous methanamine (4.7 g, 90 mmol) at 0

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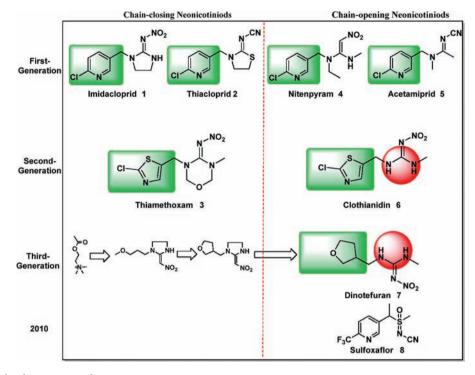


Figure 1. Commercialized neonicotinoids.

°C, and the resulting solution was stirred at room temperature for 24 h. The reaction was evaporated to dryness, and the residue was purified by chromatography over silica gel (petroleum ether/ethyl acetate, 1:1) to give 4.7 g (35.1% yield) of **22** as a yellow solid: GC-MS, m/z (%) 148 ([M]⁺, 30), 101 (100), 84 (23), 55 (96).

N-((6-Chloropyridin-3-yl)methyl)-N-ethyl-N-methyl-2-nitroethene-1,1-diamine (23). Method 1. N-((6-Chloropyridin-3-yl)methyl)-N-ethyl-N-methyl-2-nitroethene-1,1-diamine was prepared as described in the literature¹¹ in 29.5% yield: mp, 78.5–80.1 °C.

Method 2. A mixture of compound 22 (5.00 g, 34 mmol), compound 19 (5.78 g, 34 mmol), and AgOAc (5.8 g, 17 mmol) in anhydrous ethanol (30 mL) was stirred at 25 °C and monitored by TLC. After filtration, the filtrate was evaporated under reduced pressure followed by chromatography purification on silica gel (dichloromethane/ethanol, 25:1) to give 4.0 g (38.1% yield) of 23 as a yellow solid: mp, 78.1–79.6 °C.

6-(((6-Chloropyridin-3-yl)methyl)(ethyl)amino)-1-methyl-5-nitro-1,2,3,4-tetrahydropyridin-2-ol (**10a**). To a mixture of compound **23** (0.3 g, 1.1 mmol), acrylaldehyde (0.21 mL, 3.3 mol), and acetonitrile (10 mL) were added two drops of BF₃-Et₂O. The resulting solution was stirred at 40 °C for 4 h, evaporated to dryness, and purified by chromatography over silica gel (dichloromethane/acetone, 1:1) to give 0.05 g (18.0% yield) of **10a** as a pale white solid: mp, 165.1–168.2 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.28 (s, 1H), 7.77 (d, *J* = 5.4 Hz, 0.5H), 7.58 (d, *J* = 5.4 Hz, 0.5H), 7.31 (d, *J* = 8.2 Hz, 1H), 4.81 (s, 1H), 4.44–4.50 (m, 2H), 4.16 (d, *J* = 14.4 Hz, 0.5H), 4.14(d, *J* = 14.4 Hz, 0.5H), 2.96–3.26 (m, 2H), 2.96 (s, 3H), 2.70–2.88 (m, 2H), 1.92–2.05 (m, 2H), 1.16–1.26 (m, 3H); IR (KBr, cm⁻¹) 3200, 1560, 1507, 1461, 1381, 730; MS (ES+) calcd for C₁₄H₂₀³⁵ClN₄O₃ (M + H)⁺, 327 (found, 327).

N-((6-Chloropyridin-3-yl))methyl)-6-ethoxy-*N*-ethyl-1-methyl-3nitro-1,4,5,6-tetrahydropyridin-2-amine (**10b**). To a solution of compound **10a** (0.3 g, 0.9 mmol) in ethanol (10 mL) were added two drops of BF₃:Et₂O. The resulting solution was refluxed for 6 h and then evaporated to dryness. The residue was purified by chromatography over silica gel (dichloromethane/acetone, 5:1) to give 0.06 g (18.8% yield) of **10b** as a yellow solid:¹² mp, 131.1–133.7 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.27 (d, J = 2.1 Hz, 1H), 7.79 (d, J = 7.5 Hz, 0.5H), 7.64 (d, J = 7.5 Hz, 0.5H), 7.3 (d, J = 8.2 Hz, 1H), 4.35– 4.48 (m, 2H), 4.18 (d, J = 14.9 Hz, 0.5H), 4.15 (d, J = 14.9 Hz, 0.5H), 2.94–3.52 (m, 4H), 2.89 (s, 3H), 2.75 (t, J = 7.7 Hz, 2H), 1.82–2.00 (m, 2H), 1.08–1.27 (m, 6H); IR (KBr, cm⁻¹) 2891, 1559, 1469, 1380, 1281, 1246, 1249, 1096, 729; HRMS (EI+) calcd for $C_{16}H_{23}^{35}CIN_4O_3$ (M⁺), 354.1459 (found, 354.1452).

General Synthetic Procedure for 10c-e and 11a-e. A catalytic amount of BF₃·Et₂O was added to the mixture of compound 23 (0.3 g, 1.1 mmol), acrylaldehyde or crotonaldehyde (3.3 mmol), and the corresponding alcohol (10 mL), and then the mixture was heated to 40 °C. The resulting solution was stirred at 40 °C and monitored by TLC. The reaction was evaporated to dryness, and the residue was purified by chromatography over silica gel (dichloromethane/acetone, 1:1) to give the corresponding product.¹³

N-((6-Chloropyridin-3-yl)methyl)-*N*-ethyl-6-methoxy-1-methyl-3nitro-1,4,5,6-tetrahydropyridin-2-amine (**10c**): yield, 16.0%; mp, 151.1−155.2 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.30 (d, *J* = 1.9 Hz, 1H), 7.83 (d, *J* = 6.9 Hz, 0.5H), 7.62 (d, *J* = 6.9 Hz, 0.5H), 7.32 (d, *J* = 7.8 Hz, 1H), 4.26–4.49 (m, 2H), 4.09–4.23 (m, 1H), 2.94– 3.29 (m, 5H), 2.91 (s, 3H), 2.73 (m, 2H), 1.78–2.04 (m, 2H), 1.23 (m, 3H); IR (KBr, cm⁻¹) 2899, 1561, 1505, 1281, 1096, 729; HRMS (ES+) calcd for C₁₅H₂₁N₄O₃³⁵ClNa (M + Na)⁺, 363.1200 (found, 363.1196); calcd for C₁₅H₂₁N₄O₃³⁷ClNa (M + Na)⁺, 365.1170 (found, 365.1186).

N-((6-Chloropyridin-3-yl)methyl)-*N*-ethyl-1-methyl-3-nitro-6-propoxy-1,4,5,6-tetrahydropyridin-2-amine (**10d**): yield, 17.8%; mp, 110.1−114.2 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.27 (d, *J* = 1.7 Hz, 1H), 7.79 (d, *J* = 7.3 Hz, 0.5H), 7.61 (d, *J* = 7.3 Hz, 0.5H), 7.29 (d, *J* = 8.2 Hz, 1H), 4.32–4.45 (m, 2H), 4.21 (d, *J* = 14.8 Hz, 0.5H), 4.09 (d, *J* = 14.8 Hz, 0.5H), 2.86–3.51 (m, 4H), 2.86 (s, 3H), 2.72 (m, 2H), 1.79–1.99 (m, 2H), 1.54 (m, 2H), 1.19 (t, *J* = 6.9 Hz, 3H), 0.83 (t, *J* = 7.2 Hz, 3H); IR (KBr, cm⁻¹) 2900, 1561, 1507, 1458, 1381, 1281, 1248, 1096, 730; HRMS (ES+) calcd for C₁₇H₂₆N₄O₃³⁵Cl (M + H)⁺, 369.1693 (found, 369.1667); calcd for C₁₇H₂₆N₄O₃³⁷Cl (M + H)⁺, 371.1664 (found, 371.1671).

N-((6-Chloropyridin-3-yl)methyl)-*N*-ethyl-6-isopropoxy-1-methyl-3-nitro-1,4,5,6-tetrahydropyridin-2-amine (**10e**): yield, 19.8%; mp, 112.8–116.6 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.30 (s, 1H), 7.78 (d, *J* = 7.6 Hz, 0.5H), 7.67 (d, *J* = 7.6 Hz, 0.5H), 7.32 (d, *J* = 7.8 Hz, 1H), 4.40–4.46 (m, 2H), 4.17 (t, *J* = 16.2 Hz, 1H), 2.87–3.58 (m, 6H), 2.71–2.77 (m, 2H), 1.83–1.94 (m, 2H), 1.09–1.22 (m, 9H); IR (KBr, cm⁻¹) 2897, 1561, 1507, 1458, 1381, 1281, 1248, 1096, 730; HRMS

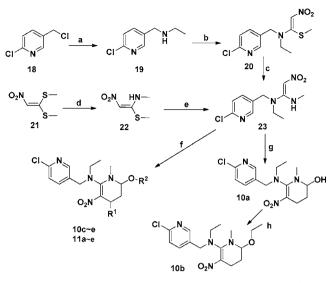
| Table 1. | Compounds | of Series | 1 | and | 2 |
|----------|-----------|-----------|---|-----|---|
|----------|-----------|-----------|---|-----|---|

| compd | R_1 | R ₂ | R ₃ | R ₄ |
|-------|--------|------------------|----------------|----------------|
| 10a | Н | Н | ethyl | methyl |
| 10b | Н | ethyl | ethyl | methyl |
| 10c | Н | methyl | ethyl | methyl |
| 10d | Н | <i>n</i> -propyl | ethyl | methyl |
| 10e | Н | isopropyl | ethyl | methyl |
| 11a | methyl | methyl | ethyl | methyl |
| 11b | methyl | ethyl | ethyl | methyl |
| 11c | methyl | <i>n</i> -propyl | ethyl | methyl |
| 11d | methyl | isopropyl | ethyl | methyl |
| 11e | methyl | allyl | ethyl | methyl |
| 12a | Н | methyl | methyl | methyl |
| 12b | Н | ethyl | methyl | methyl |
| 12c | Н | <i>n</i> -propyl | methyl | methyl |
| 12d | Н | isopropyl | methyl | methyl |
| 12e | Н | allyl | methyl | methyl |
| 13a | methyl | methyl | methyl | methyl |
| 13b | methyl | ethyl | methyl | methyl |
| 13c | methyl | <i>n</i> -propyl | methyl | methyl |
| 13d | methyl | isopropyl | methyl | methyl |
| 14a | methyl | ethyl | Н | methyl |
| 14b | methyl | <i>n</i> -propyl | Н | methyl |
| 14c | methyl | isopropyl | Н | methyl |
| 15a | ethyl | methyl | Н | methyl |
| 15b | ethyl | ethyl | Н | methyl |
| 15c | ethyl | <i>n</i> -propyl | Н | methyl |
| 15d | ethyl | isopropyl | Н | methyl |
| 16a | methyl | methyl | Н | ethyl |
| 16b | methyl | ethyl | Н | ethyl |
| 16c | methyl | <i>n</i> -propyl | Н | ethyl |
| 16d | methyl | isopropyl | Н | ethyl |
| 17a | ethyl | methyl | Н | ethyl |
| 17b | ethyl | ethyl | Н | ethyl |
| 17c | ethyl | <i>n</i> -propyl | Н | ethyl |
| 17d | ethyl | isopropyl | Н | ethyl |

(ES+) calcd for $C_{17}H_{25}N_4O_3Na^{35}Cl (M + Na)^+$, 391.1513 (found, 391.1513); calcd for $C_{17}H_{25}N_4O_3Na^{37}Cl (M + Na)^+$, 393.1483 (found, 393.1512).

N-((6-Chloropyridin-3-yl)methyl)-*N*-ethyl-6-methoxy-1,4-dimethyl-3-nitro-1,4,5,6-tetrahydropyridin-2-amine (**11a**): yield, 22.7%; mp, 125.1–129.2 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.26 (s, 1H), 7.95 (d, *J* = 5.0 Hz, 0.5H), 7.50 (d, *J* = 5.0 Hz, 0.5H), 7.29 (d, *J* = 7.8 Hz, 1H), 4.30–4.47 (m, 2H), 4.20 (m, 1H), 3.02–3.41 (m, 5H), 2.95 (s, 3H), 2.28 (m, 1H), 1.59–1.69 (m, 2H), 1.30 (t, *J* = 6.7 Hz, 3H),

Scheme 1. Preparation of Compounds 10a-11e of Series 1^a



^{*a*}Reagents and conditions: (a) ethylamine, MeCN, 0-5 °C; (b) 1,1bis(methylthio)-2-nitroethene, EtOH, refluxing; (c) methylamine, EtOH, refluxing; (d) methylamine, EtOH, refluxing; (e) compound **19**, AgOAc, EtOH, 25 °C; (f) acrylaldehyde or crotonaldehyde, corresponding alcohol, BF₃·Et₂O, 40 °C; (g) acrylaldehyde, MeCN, BF₃·Et₂O, 40 °C; (h) EtOH, BF₃·Et₂O, 40 °C.

 $0.98-1.18~(m,~3H);~IR~(KBr,~cm^{-1})~2900,~1561,~1507,~1458,~1381,~1281,~1248,~1096,~730;~HRMS~(ES+)~calcd~for~C_{16}H_{23}N_4O_3Na^{35}Cl~(M+Na)^+,~377.1356~(found,~377.1324);~calcd~for~C_{16}H_{23}N_4O_3Na^{37}Cl~(M+Na)^+,~379.1327~(found,~379.1336).$

N-((6-Chloropyridin-3-yl)methyl)-6-ethoxy-*N*-ethyl-1,4-dimethyl-3-nitro-1,4,5,6-tetrahydropyridin-2-amine (**11b**): yield, 21.5%; mp, 120.1–123.7 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.26 (s, 1H), 7.95 (d, *J* = 5.0 Hz, 0.5H), 7.50 (d, *J* = 5.0 Hz, 0.5H), 7.29 (d, *J* = 7.8 Hz, 1H), 4.24–4.48 (m, 3H), 3.24–3.53 (m, 4H), 2.95 (s, 3H), 2.25–2.29 (m, 1H), 1.56–1.68 (m, 2H), 1.28 (m, 3H), 1.15 (t, *J* = 5.8 Hz, 3H), 0.98–1.07 (m, 3H); IR (KBr, cm⁻¹) 2900, 1561, 1507, 1458, 1381, 1281, 1248, 1096, 730; HRMS (ES+) calcd for C₁₇H₂₅N₄O₃Na³⁵Cl (M + Na)⁺, 391.1513 (found, 391.1513); calcd for C₁₇H₂₅N₄O₃Na³⁷Cl (M + Na)⁺, 393.1483 (found, 393.1518).

N-((6-Chloropyridin-3-yl)methyl)-*N*-ethyl-1,4-dimethyl-3-nitro-6propoxy-1,4,5,6-tetrahydropyridin-2-amine (**11c**): yield, 22.8%; mp, 118.1–121.2 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.28 (s, 1H), 7.98 (d, *J* = 7.0 Hz, 0.5H), 7.50 (d, *J* = 7.0 Hz, 0.5H), 7.29 (d, *J* = 7.9 Hz, 1H), 4.53–4.28 (m, 3H), 3.03–3.41 (m, 4H), 2.95 (s, 3H), 2.28 (m, 1H), 1.51–1.69 (m, 2H), 1.51 (m, 2H), 1.28 (t, *J* = 8.7 Hz, 3H), 0.99–1.11 (m, 3H), 0.84 (t, *J* = 7.2 Hz, 3H); IR (KBr, cm⁻¹) 2900, 1561, 1507, 1458, 1381, 1281, 1248, 1096, 730; HRMS (ES+) calcd for

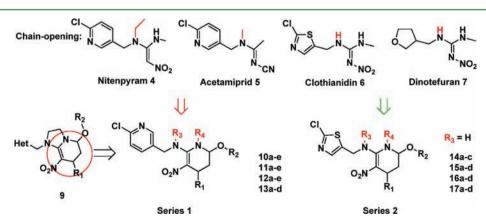


Figure 2. Chemical structures of cyclic and acyclic neonicotinoids, 9, and its acyclic analogues.

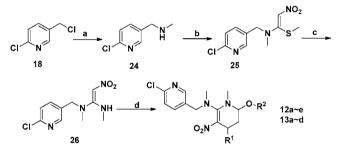
 $C_{18}H_{28}N_4O_3{}^{35}Cl~(M~+~H)^+,~383.1850$ (found, 383.1815); calcd for $C_{18}H_{28}N_4O_3{}^{37}Cl~(M~+~H)^+,~385.1820$ (found, 385.1841).

N-((6-Chloropyridin-3-yl)methyl)-*N*-ethyl-6-isopropoxy-1,4-dimethyl-3-nitro-1,4,5,6-tetrahydropyridin-2-amine (11d): yield, 23.2%; mp, 104.1−107.2 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.28 (s, 1H), 7.99 (d, *J* = 7.6 Hz, 0.5H), 7.51 (d, *J* = 7.6 Hz, 0.5H), 7.31 (d, *J* = 6.3 Hz, 1H), 4.25−4.49 (m, 3H), 3.06−3.67 (m, 3H), 2.95 (s, 3H), 2.22 (m, 1H), 1.54−1.72 (m, 2H), 1.28 (t, *J* = 6.69 Hz, 3H), 1.13 (d, *J* = 6.0 Hz, 3H), 1.07 (d, *J* = 6.1 Hz, 3H), 0.99 (d, *J* = 6.4 Hz, 3H); IR (KBr, cm⁻¹) 2896, 1561, 1507, 1458, 1381, 1281, 1248, 1096, 729; HRMS (ES+) calcd for C₁₈H₂₈N₄O₃³⁵Cl (M + H)⁺, 385.1820 (found, 383.1831); calcd for C₁₈H₂₈N₄O₃³⁷Cl (M + H)⁺, 385.1820 (found, 385.1867).

6-(Allyloxy)-N-((6-chloropyridin-3-yl)methyl)-N-ethyl-1,4-dimethyl-3-nitro-1,4,5,6-tetrahydropyridin-2-amine (**11e**): yield, 25.5%; mp, 111.9–116.7 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.26 (s, 1H), 7.96 (d, *J* = 7.2 Hz, 0.5H), 7.50 (d, *J* = 7.2 Hz, 0.5H), 7.31 (d, *J* = 8.2 Hz, 1H), 5.75–5.83 (m, 1H), 5.15–5.24 (m, 2H), 4.28–4.49 (m, 3H), 3.77–4.02 (m, 2H), 3.01–3.36 (m, 2H), 2.92 (s, 3H), 2.28–2.32 (m, 1H), 1.56–1.78 (m, 2H), 1.28 (t, *J* = 6.1 Hz, 3H); IR (KBr, cm⁻¹) 2896, 1561, 1507, 1458, 1381, 1281, 1248, 1096, 729; HRMS (ES+) calcd for C₁₈H₂₅N₄O₃Na³⁵Cl (M + Na)⁺, 403.1513 (found, 403.1520); calcd for C₁₈H₂₅N₄O₃Na³⁷Cl (M + Na)⁺, 405.1483 (found, 405.1486).

General Synthetic Procedure for 12a-e and 13a-d (Scheme 2). Compounds 24, 25, and 26 were synthesized following the same route as 19, 20, and 21.^{10,11} A catalytic amount of BF₃·Et₂O was added to

Scheme 2. Preparation of Compounds 12a-13d of Series 1^a



^{*a*}Reagents and conditions: (a) methylamine, MeCN, 0-5 °C; (b) 1,1bis(methylthio)-2-nitroethene, EtOH, refluxing; (c) methylamine, EtOH, refluxing; (d) acrylaldehyde or crotonaldehyde, corresponding alcohol, BF₃-Et₂O, 40 °C.

the mixture of compound **26** (0.3 g, 1.1 mmol), acrylaldehyde or crotonaldehyde (3.3 mmol), and the corresponding alcohol (10 mL). Then the resulting solution was stirred at 40 $^{\circ}$ C, and the progress of the reaction was monitored by TLC. The reaction mixture was concentrated by rotary evaporator, and the residue was purified by chromatography on silica gel (dichloromethane/acetone, 1:1) to give the corresponding product.

N-((6-Ĉhloropyridin-3-yl)methyl)-6-methoxy-*N*,1-dimethyl-3nitro-1,4,5,6-tetrahydropyridin-2-amine **12a::** yield, 26.9%; mp, 101.2−104.5 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.31 (s, 1H), 7.72 (m, 0.5H), 7.61 (m, 0.5H), 7.34 (d, *J* = 8.1 Hz, 1H), 4.31−4.50 (m, 2H), 4.15 (m, 1H), 3.29−3.34 (m, 3H), 2.91 (s, 3H), 2.76 (m, 5H), 1.84−2.09 (m, 2H); IR (KBr, cm⁻¹) 2900, 1561, 1508, 1458, 1380, 1281, 1248, 1096, 730; HRMS (ES+) calcd for C₁₄H₁₉N₄O₃Na³⁵Cl (M + Na)⁺, 349.1043 (found, 349.1013); calcd for C₁₄H₁₉N₄O₃Na³⁷Cl (M + Na)⁺, 351.1014 (found, 351.1017).

N-((6-Chloropyridin-3-yl)methyl)-6-ethoxy-*N*,1-dimethyl-3-nitro-1,4,5,6-tetrahydropyridin-2-amine **12b**: yield, 25.8%; mp, 94.3−98.2 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.30 (s, 1H), 7.64 (d, *J* = 7.5 Hz, 1H), 7.33 (d, *J* = 8.2 Hz, 1H), 4.35−4.39 (m, 2H), 4.14−4.17 (m, 1H), 3.38−3.49 (m, 2H), 2.88 (s, 3H), 2.76 (m, 5H), 1.84−1.87 (m, 2H), 1.17 (t, *J* = 6.9 Hz, 3H); IR (KBr, cm⁻¹) 2896, 1561, 1507, 1458, 1383, 1282, 1248, 1096, 730; HRMS (ES+) calcd for C₁₅H₂₁N₄O₃Na³⁵Cl (M + Na)⁺, 365.1170 (found, 365.1176). *N*-((6-Chloropyridin-3-yl)methyl)-*N*,1-dimethyl-3-nitro-6-propoxy-1,4,5,6-tetrahydropyridin-2-amine **12c**: yield, 21.4%; mp, 80.1–84.2 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.30 (d, *J* = 2.2 Hz, 1H), 7.63 (m, 1H), 7.33 (d, *J* = 8.2 Hz, 1H), 4.39–4.50 (m, 2H), 4.18 (m, 1H), 3.33–3.39 (m, 3H), 2.89 (s, 3H), 2.75 (m, 5H), 1.88–2.07 (m, 2H), 1.51–1.58 (m, 2H), 0.87 (t, *J* = 7.4 Hz, 3H); IR (KBr, cm⁻¹) 2896, 1561, 1507, 1458, 1381, 1281, 1248, 1096, 729; HRMS (ES+) calcd for C₁₆H₂₃N₄O₃Na³⁵Cl (M + Na)⁺, 377.1356 (found, 377.1336); calcd for C₁₆H₂₃N₄O₃Na³⁷Cl (M + Na)⁺, 379.1327 (found, 379.1295).

N-((6-Chloropyridin-3-yl)methyl)-6-isopropoxy-*N*,1-dimethyl-3nitro-1,4,5,6-tetrahydropyridin-2-amine **12d**: yield, 21.7%; mp, 85.2−89.6 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.30 (d, *J* = 2.1 Hz, 1H), 7.66 (m, 1H), 7.34 (d, *J* = 8.2 Hz, 1H), 4.36−4.49 (m, 2H), 4.18 (m, 1H), 3.64 (m, 1H), 2.88 (s, 3H), 2.75−2.79 (m, 5H), 1.92 (m, 2H), 1.15 (d, *J* = 5.7 Hz, 3H), 1.11 (d, *J* = 6.2 Hz, 3H); IR (KBr, cm⁻¹) 2896, 1562, 1507, 1458, 1381, 1281, 1248, 1096, 729; HRMS (ES+) calcd for C₁₆H₂₃N₄O₃Na³⁵Cl (M + Na)⁺, 377.1356 (found, 377.1352); calcd for C₁₆H₂₃N₄O₃Na³⁷Cl (M + Na)⁺, 379.1327 (found, 379.1326).

6-(Allyloxy)-N-((6-chloropyridin-3-yl)methyl)-N, 1-dimethyl-3nitro-1,4,5,6-tetrahydropyridin-2-amine **12e**: yield, 22.2%; mp, 76.0–80.1 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.30 (s, 1H), 7.62– 7.75 (m, 1H), 7.34 (d, *J* = 8.2 Hz, 1H), 5.83 (m, 1H), 5.19–5.25 (m, 2H), 3.93–4.55 (m, 5H), 2.89 (s, 3H), 2.78 (m, 2H), 2.75 (s, 3H), 1.86–2.10 (m, 2H); IR (KBr, cm⁻¹) 2901, 1562, 1507, 1458, 1381, 1281, 1248, 1096.34, 729; HRMS (ES+) calcd for C₁₆H₂₁N₄O₃Na³⁵Cl (M + Na)⁺, 375.1200 (found, 375.1201); calcd for C₁₆H₂₁N₄O₃Na³⁷Cl (M + Na)⁺, 377.1170 (found, 377.1175).

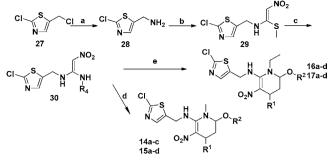
N-((6-Chloropyridin-3-yl)methyl)-6-methoxy-*N*, 1, 4-trimethyl-3nitro-1, 4, 5, 6-tetrahydropyridin-2-amine **13a**: yield, 31.6%; mp, 126.1–130.2 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.28 (s, 1H), 7.85 (m, 0.5H), 7.52 (m, 0.5H), 7.33 (d, *J* = 7.9 Hz, 1H), 4.24–4.61 (m, 3H), 3.27 (m, 3H), 2.96 (s, 3H), 2.81 (s, 3H), 2.35 (m, 1H), 1.65 (m, 2H), 1.11–1.29 (m, 3H); IR (KBr, cm⁻¹) 2896, 1562, 1507, 1459, 1381, 1281.07, 1248, 1100, 727; HRMS (ES+) calcd for $C_{15}H_{21}N_4O_3Na^{35}Cl (M + Na)^+$, 365.1170 (found, 365.1189).

N-((6-Chloropyridin-3-yl)methyl)-6-ethoxy-*N*, 1,4-trimethyl-3nitro-1,4,5,6-tetrahydropyridin-2-amine **13b**: yield, 25.1%; mp, 126.1–129.8 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.28 (s, 1H), 7.83–7.86 (m, 0.5H), 7.52–7.53 (m, 0.5H), 7.32 (d, *J* = 8.2 Hz, 1H), 4.25–4.57 (m, 3H), 3.34–3.38 (m, 2H), 2.95 (s, 3H), 2.79 (s, 3H), 2.28–2.33 (m, 1H), 1.64 (m, 2H), 1.13–1.29 (m, 6H); IR (KBr, cm⁻¹) 2896, 1562, 1507, 1459, 1381, 1281, 1248, 1096, 727; HRMS (ES+) calcd for C₁₆H₂₃N₄O₃Na³⁵Cl (M + Na)⁺, 377.1356 (found, 377.1333); calcd for C₁₆H₂₃N₄O₃Na³⁷Cl (M + Na)⁺, 379.1326 (found, 379.1327).

N-((6-Chloropyridin-3-yl)methyl)-*N*,1,4-trimethyl-3-nitro-6-propoxy-1,4,5,6-tetrahydropyridin-2-amine **13c**: yield, 32.7%; mp, 102.1−105.2 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.32 (s, 1H), 7.52−7.88 (m, 1H), 7.32 (d, *J* = 7.6 Hz, 1H), 4.30−4.59 (m, 3H), 3.25−3.39 (m, 2H), 2.95 (s, 3H), 2.80 (s, 3H), 2.30−2.35 (m, 1H), 1.63−2.09 (m, 2H), 1.51 (m, 2H), 1.14−1.29 (m, 3H), 0.84 (t, *J* = 6.9 Hz, 3H); IR (KBr, cm⁻¹) 2899, 1562, 1507, 1459, 1381, 1281, 1248, 1096, 730; HRMS (ES+) calcd for C₁₇H₂₅N₄O₃Na³⁵Cl (M + Na)⁺, 391.1513 (found, 391.1487); calcd for C₁₇H₂₅N₄O₃Na³⁷Cl (M + Na)⁺, 393.1483 (found, 393.1482).

N-((6-Chloropyridin-3-yl)methyl)-6-isopropoxy-*N*,1,4-trimethyl-3nitro-1,4,5,6-tetrahydropyridin-2-amine **13d**: yield, 30.1%; mp, 108.1−111.2 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.28 (s, 1H), 7.53−7.89 (m, 1H), 7.33 (d, *J* = 5.6 Hz, 1H), 4.34−4.80 (m, 3H), 3.26 (m, 1H), 2.94 (s, 3H), 2.79 (s, 3H), 2.24−2.29 (m, 1H), 1.28−2.04 (m, 2H), 1.10−1.26 (m, 6H), 1.06 (m, 3H); IR (KBr, cm⁻¹) 2897, 1562, 1508, 1459.79, 1381, 1281, 1248, 1096, 731; HRMS (ES+) calcd for C₁₇H₂₆N₄O₃³⁵Cl (M + H)⁺, 371.1664 (found, 371.1690).

Synthesis of Series 2 (Scheme 3). (2-Chlorothiazol-5-yl)methanamine (28). Compound 28 was prepared as described in the literature¹⁴ in 32.4% yield: GC-MS, m/z(%) 148 ([M]⁺, 15), 132 (9), 113 (100), 96 (1).



^{*a*}Reagents and conditions: (a) NH₃·H₂O, EtOH, room temperature; (b) 1,1-bis(methylthio)-2-nitroethene, EtOH, refluxing; (c) methylamine or ethylamine, EtOH, refluxing; (d) CH₂Cl₂, crotonaldehyde or *trans*-2-pentenal, corresponding alcohol, BF₃·Et₂O, 20 °C; (e) CH₂Cl₂, crotonaldehyde or *trans*-2-pentenal, corresponding alcohol, BF₃·Et₂O, 20 °C.

N-((2-Chlorothiazol-5-yl)methyl)-1-(methylthio)-2-nitroethenamine (**29**). Compound **29** was prepared as described in the literature¹⁵ in 63.8% yield: MS (ES+) calcd for $C_7H_8^{35}ClN_3O_2S_2$ (M⁺), 265 (found, 265).

N-((2-Chlorothiazol-5-yl)methyl)-N-methyl-2-nitroethene-1,1-diamine (30). Compound 30 was prepared as described in the literature.¹⁵

General Synthetic Procedure for 14a-c, 15a-d, 16a-d and 17a-d. A mixture of compound 30 (0.375 g, 1.5 mmol), crotonaldehyde or *trans*-2-pentenal (3.3 mmol), and the corresponding alcohol (5 mL) in CH₂Cl₂ (10 mL) was stirred and cooled to 20 °C, and then a catalytic amount of BF₃·Et₂O was added to the mixture. The resulting solution was stirred at 20 °C, and the progress of the reaction was monitored by TLC. The reaction was evaporated to dryness, and the residue was purified by chromatography over silica gel (dichloromethane/acetone, 12:1) to give the corresponding product.

N-((2-Chlorothiazol-5-yl)methyl)-6-ethoxy-1,4-dimethyl-3-nitro-1,4,5,6-tetrahydropyridin-2-amine **14a**: yield, 14.7%; ¹H NMR (400 MHz, CDCl₃) δ 11.05 (s, 1H), 7.49 (s, 1H), 4.59–4.62 (m, 2H), 4.45 (dd, J_1 = 4.4 Hz, J_2 = 7.6 Hz, 1H), 3.38–3.48 (m, 2H), 3.11 (s, 3H), 1.91–1.98 (m, 1H), 1.13–1.33 (m, 2H), 1.20 (t, J = 7.2 Hz, 3H), 1.15 (d, J = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 159.3, 152.5, 139.5, 136.8, 116.4, 88.4, 62.7, 42.3, 40.4, 34.2, 25.9, 17.4, 15.3.; HRMS (ES+) calcd for C₁₃H₂₀N₄O₃S³⁷Cl (M + H)⁺, 347.0945 (found, 347.0946); calcd for C₁₃H₂₀N₄O₃S³⁷Cl (M + H)⁺, 349.0915 (found, 349.0932).

N-((2-Chlorothiazol-5-yl)methyl)-1,4-dimethyl-3-nitro-6-propoxy-1,4,5,6-tetrahydropyridin-2-amine **14b**: yield, 13.9%; ¹H NMR (400 MHz, CDCl₃) δ 11.10 (s, 1H), 7.48 (s, 1H), 4.59–4.64 (m, 2H), 4.39 (s, 1H), 3.29–3.47 (m, 2H), 3.12 (s, 3H), 1.83–1.96 (m, 1H), 1.56–1.63 (m, 2H), 1.29 (d, *J* = 6.8 Hz, 3H), 1.08–1.15 (m, 2H), 0.93 (t, *J* = 7.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 159.4, 152.4, 139.4, 137.0, 116.9, 89.0, 70.7, 42.3, 38.3, 31.5, 27.4, 22.9, 18.3, 10.7; HRMS (ES+) calcd for $C_{14}H_{22}N_4O_3S^{35}Cl$ (M + H)⁺, 361.1101 (found, 361.1112); calcd for $C_{14}H_{22}N_4O_3S^{37}Cl$ (M + H)⁺, 363.1072 (found, 363.1082).

N-((2-Chlorothiazol-5-yl)methyl)-6-isopropoxy-1,4-dimethyl-3nitro-1,4,5,6-tetrahydropyridin-2-amine **14c**: yield, 14.7%; mp, 117.7−118.5 °C; ¹H NMR (400 MHz, CDCl₃) δ 11.07 (s, 1H), 7.49 (s, 1H), 4.57−4.60 (m, 2H), 4.49 (dd, J_1 = 5.2 Hz, J_2 = 6.8 Hz, 1H), 3.18−3.23 (m, 1H), 3.10 (s, 3H), 1.91−1.94 (m, 1H), 1.15−1.22 (m, 2H), 1.22 (d, J = 4 Hz, 3H), 1.18 (d, 3H), 1.6 (d, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 59.3, 152.5, 139.5, 136.8, 116.4, 88.4, 62.7, 42.3, 40.4, 34.2, 25.9, 17.4, 15.3; IR (KBr, cm⁻¹) 2962, 1573, 1524, 1354, 1357, 1276, 1172, 1059, 919, 748; HRMS (ES+) calcd for C₁₄H₂₂N₄O₃NaS³⁷Cl (M + Na)⁺, 383.0921 (found, 383.0930); calcd for C₁₄H₂₂N₄O₃NaS³⁷Cl (M + Na)⁺, 385.0891 (found, 385.0880).

N-((2-Chlorothiazol-5-yl)methyl)-4-ethyl-6-methoxy-1-methyl-3nitro-1,4,5,6-tetrahydropyridin-2-amine **15a**: yield, 15.2%; mp, 101.0–102.4 °C; ¹H NMR (400 MHz, CDCl₃) δ 11.04 (s, 1H), 7.49 (s, 1H), 4.59–4.62 (m, 2H), 4.39 (dd, J_1 = 4.8 Hz, J_2 = 8.4 Hz, 1H), 3.26 (s, 3H), 3.11 (s, 3H), 2.02–2.11 (m, 1H), 1.88–1.93 (m, 1H), 1.68–1.73 (m, 1H), 1.24–1.32 (m, 2H), 0.93 (t, J = 7.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 159.3, 152.5, 139.5, 136.8, 116.4, 89.5, 53.9, 42.3, 40.5, 32.0, 29.9, 24.3, 11.5; IR (KBr, cm⁻¹) 2962, 1595, 1409, 1335, 1212, 1134, 1049, 963, 863; HRMS (ES+) calcd for C₁₃H₂₀N₄O₃S³⁵Cl (M + H)⁺, 347.0945 (found, 347.0951); calcd for C₁₃H₂₀N₄O₃S³⁷Cl (M + H)⁺, 349.0915 (found, 349.0926).

N-((2-Chlorothiazol-5-yl)methyl)-6-ethoxy-4-ethyl-1-methyl-3nitro-1,4,5,6-tetrahydropyridin-2-amine **15b**: yield, 14.3%; mp, 103.0−103.8 °C; ¹H NMR (400 MHz, CDCl₃) δ 11.10 (s, 1H), 7.49 (s, 1H), 4.58−4.64 (m, 2H), 4.44 (dd, J_1 = 4.8 Hz, J_2 = 8.4 Hz, 1 H), 3.03−3.19 (m, 2H), 3.11 (s, 3H), 2.07−2.13 (m, 1H), 1.85−1.92 (m, 1H), 1.65−1.74 (m, 1H), 1.19−1.27 (m, 2H), 1.23 (t, J = 7.2 Hz, 3H), 0.93 (t, J = 7.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 159.4, 152.4, 139.5, 136.8, 116.3, 88.6, 62.5, 42.3, 40.3, 32.1, 30.7, 24.2, 15.3, 11.5; IR (KBr, cm⁻¹) 2970, 1585, 1524, 1409, 1335, 1276, 1146, 1059, 912, 748; HRMS (ES+) calcd for C₁₄H₂₂N₄O₃S³⁷Cl (M + H)⁺, 361.1101 (found, 361.1107); calcd for C₁₄H₂₂N₄O₃S³⁷Cl (M + H)⁺, 363.1072 (found, 363.1076).

N-((2-Chlorothiazol-5-yl)methyl)-4-ethyl-1-methyl-3-nitro-6-propoxy-1,4,5,6-tetrahydropyridin-2-amine **15c**: yield, 14.2%; mp, 89.0−91.6 °C; ¹H NMR (400 MHz, CDCl₃) δ 11.10 (s, 1H), 7.50 (s, 1H), 4.59−4.64 (m, 2H), 4.45 (dd, J_1 = 4.8 Hz, J_2 = 8.4 Hz, 1H), 3.29−3.36 (m, 2H), 3.11 (s, 3H), 2.08−2.13 (m, 1H), 1.84−1.91 (m, 1H), 1.68−1.75 (m, 1H), 1.24−1.32 (m, 2H), 0.92−0.96 (m, 8H); ¹³C NMR (100 MHz, CDCl₃) δ 159.4, 152.5, 139.5, 136.8, 116.4, 88.7, 68.6, 42.3, 40.3, 32.1, 30.6, 24.3, 23.0, 11.5, 10.6; IR (KBr, cm⁻¹) 2955, 1587, 1406, 1331, 1209, 1138, 1059, 904, 748; HRMS (ES+) calcd for C₁₅H₂₄N₄O₃S³⁵Cl (M + H)⁺, 377.1228 (found, 377.1212).

N-((2-Chlorothiazol-5-yl))methyl)-4-ethyl-6-isopropoxy-1-methyl-3-nitro-1,4,5,6-tetrahydropyridin-2-amine **15d**: yield, 13.4%; mp, 105.2−105.3 °C; ¹H NMR (400 MHz, CDCl₃) δ 11.09 (s, 1H), 7.50 (s, 1H), 4.57−4.60 (m, 2H), 4.46 (dd, J_1 = 4.8 Hz, J_2 = 8.8 Hz, 1H), 3.33−3.36 (m, 1H), 3.10 (s, 3H), 2.10−2.14 (m, 1H), 1.83−1.86 (m, 1H), 1.69−1.75 (m, 1H), 1.25−1.31 (m, 2H), 1.20 (d, J = 4 Hz, 3H), 1.18 (d, J = 4 Hz, 3H), 0.95 (t, J = 14.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 159.4, 152.4, 139.6, 136.7, 116.4, 87.3, 70.4, 42.3, 40.1, 32.2, 32.1, 25.4, 24.2, 22.8, 11.6; IR (KBr, cm⁻¹) 2962, 1613, 1509, 1409, 1357, 1224, 1149, 1045, 919, 748; HRMS (ES+) calcd for C₁₅H₂₃³⁵ClN₄NaO₃S (M + Na)⁺, 397.1077 (found, 397.1064); calcd for C₁₅H₂₃³⁷ClN₄NaO₃S (M + Na)⁺, 399.1048 (found, 399.1020).

N-((2-Chlorothiazol-5-yl)methyl)-1-ethyl-6-methoxy-4-methyl-3nitro-1,4,5,6-tetrahydropyridin-2-amine **16a**: yield, 15.9%; mp, 150.4−151.3 °C; ¹H NMR (400 MHz, CDCl₃) δ 10.65 (s, 1H), 7.49 (s, 1H), 4.58 (s, 2H), 4.38 (dd, J_1 = 4.8 Hz, J_2 = 7.2 Hz, 1H), 3.35−3.42 (m, 2H), 3.28 (s, 3H), 1.98−2.05 (m, 1H), 1.39 (t, J = 7.2 Hz, 3H), 1.27−1.31 (m, 1H), 1.09−1.17 (m, 1H), 1.00 (d, J = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 158.3, 141.4, 134.1, 117.8, 85.1, 53.8, 50.9, 46.5, 42.1, 33.4, 25.5, 17.7, 15.9; IR (KBr, cm⁻¹) 2962.96, 1584.22, 1509.98, 1432.02, 1413.46, 1205.57, 1779.58, 1038.52, 942.00, 748.96; HRMS (ES+) calcd for C₁₃H₁₉³⁵ClN₄NaO₃S (M + Na)⁺, 369.0764 (found, 369.0758); calcd for C₁₃H₁₉³⁷ClN₄NaO₃S (M + Na)⁺, 371.0735 (found, 371.0719).

N-((2-Chlorothiazol-5-yl)methyl)-6-ethoxy-1-ethyl-4-methyl-3nitro-1,4,5,6-tetrahydropyridin-2-amine **16b**: yield, 15.5%; mp, 94.1−94.4 °C; ¹H NMR (400 MHz, CDCl₃) δ 11.14 (s, 1H), 7.46 (s, 1H), 4.52−4.69 (m, 2H), 4.52−4.69 (m, 1H), 3.48−3.53 (m, 2H), 3.33−3.38 (m, 2H), 1.93−1.97 (m, 1H), 1.38 (t, *J* = 7.2 Hz, 3H), 1.31 (d, *J* = 6.8 Hz, 3H), 1.20−1.29 (m, 2H), 1.25 (t, *J* = 7.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 158.3, 153.1, 140.4, 135.7, 87.3, 63.8, 46.6, 42.0, 31.3, 27.0, 18.5, 15.8, 15.1; IR (KBr, cm⁻¹) 2977, 1591, 1532, 1458, 1368, 1276, 1179, 1049, 949, 774; HRMS (ES+) calcd for C₁₄H₂₂N₄O₃S³⁷Cl (M + H)⁺, 361.1101 (found, 361.1103); calcd for C₁₄H₂₂N₄O₃S³⁷Cl (M + H)⁺, 363.1072 (found, 363.1072).

N-((2-Chlorothiazol-5-yl)methyl)-1-ethyl-4-methyl-3-nitro-6-propoxy-1,4,5,6-tetrahydropyridin-2-amine **16c**: yield, 15.0%; ¹H NMR (400 MHz, CDCl₃) δ 11.13 (s, 1H), 7.46 (s, 1H), 4.55–4.69 (m, 2H),

4.51 (t, J = 2.8 Hz, 1H), 3.42 (t, J = 6.4 Hz, 2H), 3.33–3.38 (m, 2H), 1.93–1.97 (m, 1H), 1.59–1.67 (m, 2H), 1.38 (t, J = 7.2 Hz, 3H), 1.31 (d, J = 6.8 Hz, 3H), 1.20–1.29 (m, 2H), 0.95 (t, J = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 158.3, 153.0, 140.4, 135.6, 117.1, 87.5, 70.2, 46.7, 42.0, 31.3, 27.0, 18.5, 22.9, 18.6, 15.8, 10.8; HRMS (ES+) calcd for C₁₅H₂₄N₄O₃S³⁵Cl (M + H)⁺, 375.1258 (found, 375.1256); calcd for C₁₅H₂₄N₄O₃S³⁷Cl (M + H)⁺, 377.1228 (found, 377.1268).

N-((2-Chlorothiazol-5-yl)methyl)-1-ethyl-6-isopropoxy-4-methyl-3-nitro-1,4,5,6-tetrahydropyridin-2-amine **16d**: yield, 13.8%; ¹H NMR (400 MHz, CDCl₃) δ 11.72 (s, 1H), 7.47 (s, 1H), 4.57 (s, 2H), 4.50 (dd, J_1 = 4.0 Hz, J_2 = 6.8 Hz, 1H), 3.69–3.75 (m, 1H), 3.36–3.41 (m, 2H), 1.95–2.00 (m, 1H), 1.91–1.93 (m, 2H), 1.72– 1.79 (m, 2H), 1.38 (t, J = 7.2 Hz, 3H), 1.17 (d, J = 6.0 Hz, 3H), 1.14 (d, J = 6.0 Hz, 3H), 1.00 (d, J = 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 158.5, 153.2, 141.1, 134.5, 117.9, 82.7, 69.0, 46.3, 42.0, 31.5, 27.2, 23.0, 18.6, 15.9, 14.1; HRMS (ES+) calcd for C₁₅H₂₄N₄O₃S³⁵Cl (M + H)⁺, 375.1258 (found, 375.1255); calcd for C₁₅H₂₄N₄O₃S³⁷Cl (M + H)⁺, 377.1228 (found, 377.1230).

N-((2-Chlorothiazol-5-yl)methyl)-1,4-diethyl-6-methoxy-3-nitro-1,4,5,6-tetrahydropyridin-2-amine **17a**: yield, 14.6%; mp, 117.1– 119.5 °C; ¹H NMR (400 MHz, CDCl₃) δ 10.68 (s, 1H), 7.48 (s, 1H), 4.59 (s, 2H), 4.38 (t, *J* = 6.0 Hz, 1H), 3.25–3.30 (m, 2H), 3.28 (s, 3H), 1.92–1.95 (m, 1H), 1.39 (t, *J* = 7.2 Hz, 3H), 1.20–1.32 (m, 2H), 0.96–1.08 (m, 2H), 0.83 (t, *J* = 7.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 158.5, 153.3, 141.4, 133.9, 117.2, 85.0, 53.5, 46.2, 42.1, 31.7, 29.8, 24.5, 15.9, 11.3; IR (KBr, cm⁻¹) 2955, 1584, 1509, 1450, 1376, 1261, 1187, 967, 748; HRMS (ES+) calcd for C₁₄H₂₁N₄O₃S³⁵ClNa (M + Na)⁺, 383.0921 (found, 383.0913); calcd for C₁₄H₂₁N₄O₃S³⁷ClNa (M + Na)⁺, 385.0891 (found, 385.0879).

N-((2-Chlorothiazol-5-yl)methyl)-6-ethoxy-1,4-diethyl-3-nitro-1,4,5,6-tetrahydropyridin-2-amine **17b**: yield, 17.0%; mp, 103.1–103.9 °C; ¹H NMR (400 MHz, CDCl₃) δ 10.73 (s, 1H), 7.46 (s, 1H), 4.59 (s, 2H), 4.43 (dd, J_1 = 5.6 Hz, J_2 = 7.2 Hz, 1H), 3.40–3.51 (m, 2H), 3.32–3.40 (m, 2H), 1.90–1.94 (m, 1H), 1.37 (t, J = 7.2 Hz, 3H), 1.21 (t, J = 7.2 Hz, 3H), 1.14–1.29 (m, 2H), 0.93–1.06 (m, 2H), 0.82 (t, J = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 158.6, 153.2, 141.3, 134.1, 117.1, 84.0, 61.9, 46.0, 42.2, 31.8, 30.4, 24.5, 15.9, 15.3, 11.3; IR (KBr, cm⁻¹) 2970.37, 1584.22, 1521.11, 1446.87, 1342.92, 1187.01, 1049.65, 956.84, 745.24; HRMS (ES+) calcd for C₁₅H₂₄N₄O₃S³⁷Cl (M + H)⁺, 377.1228 (found, 377.1241).

N-((2-Chlorothiazol-5-yl)methyl)-1,4-diethyl-3-nitro-6-propoxy-1,4,5,6-tetrahydropyridin-2-amine **17c**: yield, 13.8%; mp, 90.2– 91.5 °C; ¹H NMR (400 MHz, CDCl₃) δ 10.72 (s, 1H), 7.47 (s, 1H), 4.59 (s, 2H), 4.44 (dd, J_1 = 5.2 Hz, J_2 = 7.2 Hz, 1H), 3.36 (t, J = 6.8 Hz, 2H), 3.27–3.33 (m, 2H), 1.91–1.96 (m, 1H), 1.58–1.62 (m, 2H), 1.38 (t, J = 7.2 Hz, 3H), 1.20–1.33 (m, 2H), 0.98–1.07 (m, 2H), 0.94 (t, J = 7.2 Hz, 3H), 0.83 (t, J = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 158.6, 153.2, 141.3, 134.1, 117.1, 84.0, 67.9, 46.0, 42.1, 31.8, 30.4, 24.5, 23.0, 15.9, 11.3, 10.7; IR (KBr, cm⁻¹) 2955, 1587, 1517, 1342, 1298, 1227, 1135, 1038, 956, 748; HRMS (ES+) calcd for C₁₆H₂₆N₄O₃S³⁵Cl (M + H)⁺, 391.1385 (found, 391.1373). *N*-((2-Chlorothiazol-5-yl)methyl)-1,4-diethyl-6-isopropoxy-3-

N-((2-*c*hlorothiazoi-5-yl)methyl)-1,4-diethyl-6-isopropoxy-3nitro-1,4,5,6-tetrahydropyridin-2-amine **17d**: yield, 13.5%; ¹H NMR (400 MHz, CDCl₃) δ 10.73 (s, 1H), 7.46 (s, 1H), 4.58 (s, 2H), 4.48 (dd, J_1 = 5.2 Hz, J_2 = 8.4 Hz, 1H), 3.34–3.42 (m, 2H), 3.24–3.27 (m, 1H), 1.89–1.95 (m, 1H), 1.38 (t, J = 7.2 Hz, 3H), 1.18 (d, J = 6.4 Hz, 3H), 1.15 (d, J = 6.0 Hz, 3H), 1.20–1.33 (m, 2H), 0.94–1.07 (m, 2H), 0.83 (t, J = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 158.6, 153.2, 141.1, 134.4, 117.2, 82.8, 69.0, 46.0, 42.0, 31.9, 30.0, 27.2, 24.5, 23.0, 15.9, 11.3; HRMS (ES+) calcd for C₁₆H₂₆N₄O₃S³⁷Cl (M + H)⁺, 391.1385 (found, 391.1394).

Biological Assay. According to statistical requirements, the bioassay was repeated three times at 25 ± 1 °C. All compounds were dissolved in *N*,*N*-dimethylformamide (AP, Shanghai Chemical Reagent Co., Ltd., Shanghai, China) and diluted with water containing Triton X-100 (0.1 mg L⁻¹) to obtain series concentrations of 500.0, 250.0, and 125.0 mg L⁻¹ and others for bioassays.

Insecticidal Test for Cowpea Aphid (*Aphis craccivora*). The insecticidal activities of title compounds against cowpea aphid (*A. craccivora*) were tested according to the previously reported procedure.^{5,16} Horsebean seedlings with 40–60 healthy apterous adults were dipped in diluted solutions of the chemicals containing Triton X-100 (0.1 mg L⁻¹) for 5 s, and then the shoots were placed in a conditioned room (25 ± 1 °C, 50% relative humidity (RH)). Water containing Triton X-100 (0.1 mg L⁻¹) was used as control. The mortality rates were assessed after 24 h. Each treatment had three repetitions, and the data were corrected and subjected to probit analysis.

Bioassay on the Repellent Effects on Cowpea Aphid (A. craccivora). To understand the responses of aphid to title compounds, a Y-tape selective olfactometer was used for examination in the laboratory.¹⁷ Horsebean seedlings without aphids were dipped in diluted solution of the chemicals containing Triton X-100 (0.1 mg L^{-1}) for 5 s and then allowed to dry. The treated horsebean seedlings were put in one arm of the olfactometer, and the control ones were put in the other side arm. Apterous adult aphids were transferred into the third arm by brush. The olfactometer was covered with black cloth in the conditioned room (25 ± 1 °C, 50% RH). The number of adults in each arm was counted 1 h later. Each treatment had three repetitions, and the data were analyzed by ANOVA.

Insecticidal Test for Armyworm (*Pseudaletia separata* Walker). The insecticidal activities of title compounds against armyworm (*P. separata* Walker) were tested according to a previously reported procedure¹⁸ by foliar application. Individual corn (*Zea mays*) leaves were placed on moistened pieces of filter paper in Petri dishes. The leaves were sprayed with the solution containing compounds and allowed to dry. The dishes were infested with 10 second-instar larvae and then placed in a conditioned room (25 ± 1 °C, 50% RH); 48 h later, the mortality rates were assessed. Each treatment had three repetitions, and the data were corrected and subjected to probit analysis as before.

RESULTS AND DISCUSSION

Synthesis. Two methods were tried to synthesize the intermediate **23**. Using method 1, the reaction was carried out at 0 °C and the yield was 29.5%. By comparison with method 1, method 2 had a higher yield of 38.1%. However, a precipitate of Ag_2S formed in the reaction, which needed additional workup. Thus, method 1 was chosen to synthesize **23**.

When the target compounds were synthesized, the reaction was very complicated and there were many byproducts, which resulted in <5% yield. To optimize this reaction, several kinds of catalysts, including protonic acid and Lewis acid, such as AlCl₃, HCl, BF₃, CH₃COOH, and H₂SO₄, were studied. In view of increasing yield (10%) and simplicity of workup, BF₃·Et₂O was used as catalyst. The solvent of this reaction was also optimized; various solvents such as polar and nonpolar ones were employed, including MeCN, THF, DMF, alcohols, and dichloromethane. The result showed that dichloromethane was suitable to carry out the reaction. Furthermore, the reaction temperature was the vital factor for the synthesis of titled compounds. The reaction temperature was varied from 0 to 40 °C; it was observed that temperatures over 25 °C would lead to the formation of much more corresponding hydroxyl, eliminating byproducts, namely, 1,4-dihydropyridine analogues. The reaction temperature must be controlled below 25 °C.

After optimization, the factors in this reaction, such as temperature, catalyst, and solvent, were fixed and gave relatively higher yield. For compounds 10a-13d, the yield reached 16.0-32.7%, whereas the yields of 14a-17d were 13.4-17.0%. Compared to the synthesis of compounds 10a-13d, the synthesis of compounds 10a-13d, the synthesis of compounds 14a-17d was more difficult. Although an improvement of reaction time and temperature was tried

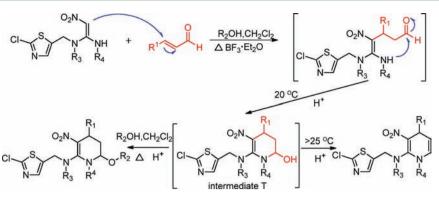


Figure 3. Mechanism to synthesize series 2.

and one-pot multicomponent reaction was also used to increase the yield and simplify the process, and even microwave synthesis was also carried out, it is still hard to get a higher yield during the synthesis of chain-opening nitromethylene neonicotinoids with cis configuration.

On the basis of our synthesis experience of this reaction, the main reason for very low yield was supposed. As shown in Figure 3, it was observed that the reaction not only for compounds 10a-13d but also for 14a-17d finished with high conversion monitored by TLC, but many spots of the product were observed on the TLC plate, which almost became a long line. A relatively low yield was obtained due to the accompanying side reactions of Michael addition, cyclization reaction, elimination reaction, and etherification. Several byproducts containing intermediates T and hydroxyl-eliminating products were isolated by column chromatography, and their structures were confirmed. Especially, the hydroxyleliminating compounds were the main products in the reaction. In addition, the conversion rate of the etherification reaction was also very low. Therefore, it was difficult to synthesize and separate target compounds. How to decrease the hydroxyl elimination reaction was the key problem of synthesis. Further optimization is in progress.

Biological Activities. Compounds 10a-13d in series 1 exhibited rather poor insecticidal activity against cowpea aphid (*A. craccivora*) at 500 mg L⁻¹. The insecticidal activities are shown in Table 2, with 11b showing the highest insecticidal activity of 31.8%. Compared with cyclic counterparts, these chain-opening molecules in series 1 all exhibited much reduced

Table 2. Insecticidal Activities of Compounds 10a-13d against Cowpea Aphid (*Aphis craccivora*)

| compd | A. craccivora mortality (%) at 500 mg L^{-1} | compd | A. craccivora mortality (%) at 500 mg L^{-1} |
|-------|--|-------|--|
| 10a | 16.2 | 12a | 21.6 |
| 10b | 17.7 | 12b | 23.3 |
| 10c | 24.3 | 12c | 15.2 |
| 10d | 23.7 | 12d | 17.6 |
| 10e | 15.3 | 12e | 15.8 |
| 11a | 27.6 | 13a | 23.4 |
| 11b | 31.8 | 13b | 17.1 |
| 11c | 27.2 | 13c | 28.3 |
| 11d | 25.8 | 13d | 25.9 |
| 11e | 19.5 | | |

insecticidal activity against cowpea aphid.⁹ Interestingly, we noted that the cowpea aphids fell down and crawled far away

from the horsebean seedling after 24 h. This observation initiated our interest in investigating the repellent effects of these compounds. We chose compound **11b** as representative and found that its repellent percentage to apterous adults was $66.4 \pm 8.6\%$, which is very interesting. There is no report of the repellent activities of known neonicotinoids such as nitenpyram, which has the same structure scaffold and does not have cis configuration present in compound **10–13**. Studies on the mechanism of repellent effects of this series of compounds are currently under investigation. This work would be helpful for the discovery of new leading compounds.

The insecticidal activities of compounds 14a-17d of series 2 against cowpea aphid (A. craccivora) are shown in Table 3. Compounds 14b, 14c, and 15c showed improved insecticidal activity against cowpea aphid with LC50 values of 0.05681, 0.08422, and 0.03855 mmol L^{-1} , respectively, compared to Paichongding, a commercial cis configuration cyclic compound with an LC_{50} of 0.09176 mmol L^{-1} (Table 3).¹⁹ The bioactivities increased by 1.1-2.4-fold. Moreover, compared to other cyclic compounds with a pyridine (PYTP)¹⁹ or thiazole ring (TZTP), with LC_{50} values of 0.26218 and 0.19001 mmol L^{-1} , respectively, the bioactivities of chain-opening series 2 also increased. Although the bioactivities of series 2 against cowpea aphid were lower than that of imidacloprid, the series 2 compounds could display better bioactivities against resistant strain brown planthopper similar to other known cis configuration analogues, such as Paichongding.

Aliphatic and aromatic substituents were available for R_3 and R_4 . On the basis of the study of Akayama and co-workers, aliphatic substituted compounds exhibited higher insecticidal activities than those bearing an aromatic group.²⁰ Therefore, aromatic substituents were not chosen in this work.

The structural difference between compounds in series 1 (10a-13d) and 2 (14a-17d) was the substituent R₃. When R₃ is a hydrogen atom as in series 2, compounds exhibited better activities than those with a methyl or ethyl group as in series 1. It suggested that this position is important for the increase of bioactivity of chain-opening cis configuration neonicotinoids compounds.

At the same time, we also found that its substituent effect was different from that in the structure of commercialized nitenpyram. The effect of substituent of nitenpyram showed that methyl and ethyl groups have good bioactivities, with an LC_{95} value of 40 ppm against green rice leafhopper planthopper, whereas replacing those groups with a hydrogen atom decreases bioactivities, giving an LC_{95} value of >200 ppm.²⁰ When R₃ changed from a hydrogen atom to methyl and then ethyl, the larger the substituent was, the lower the

| Table 3. Insecticidal Activities of Compounds 14a-c, 15a-d, |
|---|
| 16b-d, and 17a-d against Cowpea Aphid (Aphis craccivora) |

| , 8 | Aphis craccivora | | | |
|--------------|---------------------------|------------------|--|--|
| Compd. | mortality | LC ₅₀ | | |
| | (%) 500mg L ⁻¹ | $(mmol L^{-1})$ | | |
| 14a | 93.2 | 0.16218 | | |
| 14b | 100 | 0.05681 | | |
| 14c | 100 | 0.08422 | | |
| 15a | 97.4 | 0.12188 | | |
| 15b | 96.4 | 0.10578 | | |
| 15c | 88.5 | 0.03855 | | |
| 15d | 83.6 | nt | | |
| 16b | 85.2 | nt | | |
| 16c | 94.6 | nt | | |
| 16d | 75 | nt | | |
| 17a | 84.5 | nt | | |
| 17b | 84 | 0.15298 | | |
| 17c | 93.4 | nt | | |
| 17d | 84.3 | 0.13882 | | |
| Imidacloprid | 100 | 0.03090 | | |
| | 100 | 0.09176 | | |
| Paichongding | Paichongding | | | |
| | 100 | 0.26218 | | |
| РҮТР | | | | |
| | 100 | 0.19001 | | |
| ТΖТР | | | | |

bioactivity was. Substituents on the nitrogen atom of the tetrahydropyridine ring (R_4) were important for improving activities of compounds 14a–17d, and different substituents resulted in distinct difference of bioactivities. Especially, the bioactivity with the methyl group was better than that with the ethyl group. It also suggested that the sterics of R_3 and R_4 are likely playing a role in enhancing bioactivity. More analogues will be synthesized to further evaluate the substituent effect.

As for the substituent R_2 , an *n*-propyl group led to slightly higher activity than an *iso*-propyl group did, whereas both are much better than methyl or ethyl analogues as exhibited by compounds **14a–c**. Compounds **14b** and **14c** displayed comparable LC_{50} values of f 0.5681 and 0.8422 mmol L^{-1} , respectively, which are considerably higher than that of compound **14a**. To further support this observation, other compounds with *n*-propyl at R_2 including compounds **15–17** all exhibit higher mortality, which displayed the same rule as the ring-closing tetrahydropyridine compounds.¹⁹ However, compared with substituents R_4 , the influence of R_2 on bioactivities is less profound.

Because some of our nitromethylene neonicotinoids with fixed cis configuration in previous work showed good insecticidal activities against army worm, the insecticidal activity of compounds 14a-c against armyworm (*P. separata* Walker) at 500 mg L⁻¹ was also measured. Mortalities of these three compounds were all 100% with LC₅₀ values at 0.40864, 0.21781, and 0.25702 mmol L⁻¹, respectively. Although the bioactivities were not as high as those of other cis configuration neonicotiniods,^{21,22} they are still promising candidates for further modifications against army worm (*P. separata* Walker).

In conclusion, two series of chain-opening nitromethylene neonicotinoids with cis configuration were designed and synthesized. Compared with cyclic ones, they showed obvious differences. The bioassays showed that some of the compounds exhibited unique repellent effects and other compounds showed better insecticidal activities against cowpea aphid and armyworm than cyclic compounds. Compound **15c** exhibited the highest insecticidal activity against cowpea aphid among the synthesized compounds; the LC_{50} value was 0.03855 mmol L^{-1} . Further studies on the particular biological behavior of compounds **10a–13d** are underway.

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